

Crystallographic Analyses of the Runx-1/AML1 (Runt domain) -DNA Interaction and Its Allosteric Control by CBFβ

The promoter regions of most genes contain binding sites for multiple transcription factors whose interactions are essential for the correct regulation of transcription. Such interactions include those between different transcription factors as well as those between subunits of the same factor. In the case of some heterodimeric factors (*e.g.*, Jun/Fos and Myc/Max) both subunits bind to specific DNA sequences, and recent X-ray studies of this type of protein have clarified the molecular mechanisms of



Fig. 1. Overviews of the CBF α - β -C/EBP β -DNA quaternary complex from three perspectives [1]. Views (**a**) from the front, (**b**) from the side and (**c**) from the top. Within CBF α , β strands and loops are depicted as red arrows and pink tubes, respectively; within CBF β , α helices, β strands and loops are depicted as green ribbons, blue arrows and cyan tubes, respectively. The Cterminal region of the C/EBP β homodimer containing the bZip domain is shown as yellow ribbons. Areas I and II are marked with circles.

their DNA binding (e.g., the "scissors grip" by the Fos-Jun heterodimer). On the other hand, some transcription factors consist of one subunit that binds to DNA and one that does not. In most cases of this type of protein, it remains unknown how a non DNA-binding subunit contributes to DNA binding and whether the mechanism by which a non-DNA-binding subunit stimulates DNA binding is shared with different transcription factors. In this type, very little structural information is available to date, *e.g.* the GABP α -GABP β -DNA complex (GABP: GA-binding protein). One well-studied example of this type of transcription factor is the core binding factor (CBF), which functions as a heterodimer consisting of a DNA-binding α subunit and a non-binding β subunit.

The CBF heterodimeric transcription factors comprised of AML/CBFA/PEBP2a/Runx and $CBF\beta/PEBP2\beta$ subunits are essential for differentiation of hematopoietic and bone cells, and their mutation is intimately related to the development of acute leukemia and cleidocranial dysplasia; (AML: acute myelogenous leukemia, CBFA: core-binding factor A and PEBP: polyomavirus enhancer-binding protein). Here we present the crystal structures of the AML1/Runx-1/CBFa(Runt domain)-CBF β (core domain)-C/EBP β (bZip)-DNA, AML1/Runx-1/CBFα(Runt domain)-C/EBPβ(bZip)-DNA, and AML1/Runx-1/CBFa(Runt domain)-DNA complexes, which are abbreviated to CBF α - β -C/ EBP β -DNA, CBF α -C/EBP β -DNA and CBF α -DNA respectively, obtained from the diffraction experiments at beamlines BL41XU and BL45XU [1]. Our focus is on the recognition of specific DNA sequences by CBF α and on the mechanism by which CBF β facilitates DNA binding.

The structure of a complex comprising the CBF α Runt domain and the CBF β core domain bound to a 26-bp DNA fragment from the CSF-1R promoter, together with the C/EBP β basic leucine zipper region homodimer, is shown in Fig. 1(a-c). Note that the structures of DNA-bound CBF α are





Fig. 2. $CBF\alpha$ - $CBF\beta$ heterodimer structure in complex with the DNA [1]. Two predominantly hydrophilic interaction areas between $CBF\alpha$ and $CBF\beta$ (Areas I and II) are marked with dotted circles.

very similar, with or without CBF β : the r.m.s. deviation of the α carbon atoms was only about 0.28 Å. Both the CBF α Runt domain and the CBF β core domain fold principally as β barrel architectures, with the structure of the former being classified as an s-type immunoglobulin fold. The Runt domain recognizes specific bases (TGTGGTT) in both the major and minor grooves of the DNA, mainly using loops. The CBF β core domain interacts with the CBF α Runt

domain at a site distant from the protein-DNA interface. The CBF α -CBF β interface extends linearly from the DNA and can be divided into two predominantly hydrophilic interaction areas – one proximal to DNA (Area I) and one distal (Area II) – with an intervening hydrophobic area (Figs. 2 and 1b). Contrary to earlier protein-protein and protein-DNA binding assays [2,3], analyses of crystals of the CBF α - β -C/ EBP β -DNA and CBF α -C/EBP β -DNA complexes revealed no direct interaction between the C/EBP β basic leucine zipper domain and the CBF α Runt domain or the CBF β core domain. CBF α -DNA binding is enhanced several folds in the presence of CBF β . To examine the regulation of CBF α -DNA binding by CBF β , we compared the structure of the CBF α - β -C/EBP β -DNA complex with those of the CBF α -C/EBP β -DNA and CBF α -DNA complexes, which lack the CBF β subunit. Somewhat surprisingly, CBF β binding causes no dramatic structural changes in the CBF α Runt domain, which implies that CBF β does not alter the



Fig. 3. Schematic representation of the stabilization network between $CBF\beta$ (cyan), $CBF\alpha$ (pink) and DNA (gray) [1].

19



overall conformation of the CBF α Runt domain, but only stabilizes it in a conformation suitable for DNA binding. As described above, CBF β interacts with CBF α via two predominantly hydrophilic regions– Areas I and II, and their stabilization affects the conformations of the flexible loops in contact with the DNA to grip the DNA molecule at both the major and minor grooves (Fig. 3).

Acute leukemia and related diseases associated with biallelic and heterozygous point mutations of AML1/Runx-1 have been reported. In the case of biallelic mutations, no functional AML1/Runx-1 allele remains; in the case of heterozygotes, the DNA binding activity of AML1/Runx-1 from one allele is impaired, with AML1/Runx-1-CBF β heterodimerization activity retained. Based on the three-dimensional structure of the CBF α -CBF β heterodimer bound to the DNA, point mutation sites in the heterozygous cases are mapped exclusively to regions of the CBF α -DNA interface (Fig. 4). It is readily apparent that these mutations impair the capacity of CBF α to bind to DNA without affecting the overall folding architecture of CBF α or CBF α -CBF β heterodimerization, which could explain their dominant negative behavior.

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Acute Myelogenous Leukemia and related diseases



Fig. 4. Structural maps of CBF α point mutations occurring in acute myelogenous leukemia and related diseases [1]. The mutated residues are shown in green ball and stick presentations. The peptide backbones of CBF α and CBF β are depicted as pink and cyan tubes, respectively.

References

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